

List of the developed methods for the detection of Drugs in PARC

No	Drug	Matrix	Sample preparation method	Method	Linear range	LOD	Detection range	Researcher	Ref
1	Carvedilol	Plasma	Dispersive liquid–liquid microextraction based on solidification of floating organic droplet	Spectrofluorimetry	40 – 300 ng mL ⁻¹	18 ng mL ⁻¹	40 – 223 ng mL ⁻¹	M.Z	(1)
2	Metoprolol Propranolol Carvedilol Diltiazem Verapamil	Plasma	Dispersive liquid–liquid microextraction	HPLC-UV	0.02 – 1 µg mL ⁻¹	0.002 µg mL ⁻¹ 0.005 µg mL ⁻¹ 0.006 µg mL ⁻¹ 0.003 µg mL ⁻¹ 0.003 µg mL ⁻¹	25.8-92.3 ng mL ⁻¹ 70.6 ng mL ⁻¹ 23.6 ng mL ⁻¹	R.F	(2)
3	Carvedilol Propranolol	Urine	Vortex-assisted liquid–liquid extraction	CE-DAD	0.005-1 µg mL ⁻¹	0.001 µg mL ⁻¹ 0.0007 µg mL ⁻¹	22 – 163 ng mL ⁻¹	R.F	(3)
4	Propranolol Metoprolol Carvedilol	Urine	Salt-assisted liquid–liquid extraction	CE-DAD	0.025-1 µg mL ⁻¹	0.008 µg mL ⁻¹ 0.005 µg mL ⁻¹ 0.015 µg mL ⁻¹	67-150 ng mL ⁻¹	R.F	(4)
5	Carbamazepine	Plasma	Stir bar sorptive extractionMagnetic solid phase extraction	HPLC-UV	0.2-12 µg mL ⁻¹ 0.05-12 µg mL ⁻¹	0.01 µg mL ⁻¹ 0.0043 µg mL ⁻¹	2.3-5.5 µg mL ⁻¹	S.A	(5)
6	Verapamil	Exhaled breath condensate	-	Spectrofluorimetry	0.02-12 µg mL ⁻¹	0.008 µg mL ⁻¹	0.059-0.067 µg mL ⁻¹	F.P	(6)
7	Verapamil	Exhaled breath condensate	Dispersive liquid–liquid microextraction	HPLC-UV	0.07-0.8 µg mL ⁻¹	-	0.07-0.09 µg mL ⁻¹	F.P	(6)

8	Phenobarbital Carbamazepine	Urine	Homogenous liquid-liquid extraction coupled with dispersive liquid-liquid microextraction	GC-FID	$0.06\text{-}100 \mu\text{g mL}^{-1}$ $0.04\text{-}100 \mu\text{g mL}^{-1}$	$0.017 \mu\text{g mL}^{-1}$ $0.010 \mu\text{g mL}^{-1}$	-	B.F	(7)
9	Ethanol	Exhaled breath condensate	-	Spectrophotometry	$300\text{-}1500 \mu\text{g mL}^{-1}$ $1600\text{-}8000 \mu\text{g mL}^{-1}$	$82.5 \mu\text{g mL}^{-1}$ $330 \mu\text{g mL}^{-1}$	$200\text{-}800 \mu\text{g mL}^{-1}$	F.P	(8)
10	Lamotrigine	Exhaled breath condensate	-	Spectrophotometry	$0.02\text{-}1 \mu\text{g mL}^{-1}$	5 ng mL^{-1}	$0.592\text{-}0.721 \mu\text{g mL}^{-1}$	A.S	(9)
11	Lamotrigine	Plasma	-	Spectrofluorimetry	$0.5\text{-}6.0 \mu\text{g mL}^{-1}$	$0.3 \mu\text{g mL}^{-1}$	$1.69\text{-}3.71 \mu\text{g mL}^{-1}$	A.S	(9)
12	Phenytoin	Plasma	-	Spectrophotometry	$67\text{-}670 \text{ ng mL}^{-1}$	21 ng mL^{-1}	$2.06\text{-}4.84 \mu\text{g mL}^{-1}$	A.S	(10)
13	Phenytoin	Exhaled breath condensate	-	Spectrophotometry	$25\text{-}450 \mu\text{g L}^{-1}$	$10 \mu\text{g L}^{-1}$	-	A.S	(10)
14	Doxorubicin	Exhaled breath condensate	-	Spectrophotometry	$20\text{-}200 \mu\text{g L}^{-1}$	$4.16 \mu\text{g L}^{-1}$	-	A.S	(11)
15	P-Cresol	Plasma	In situ surfactant -based solid phase microextraction	HPLC-FL	$0.5\text{-}8 \mu\text{g mL}^{-1}$	$0.324 \mu\text{g mL}^{-1}$	-	A.S	(12)
16	Phenobarbital Phenytoin Carbamazepine Carbamazepine epoxide	Plasma	-	HPLC-UV	$1\text{-}40 \mu\text{g mL}^{-1}$ $1\text{-}30 \mu\text{g mL}^{-1}$ $0.3\text{-}15 \mu\text{g mL}^{-1}$ $0.5\text{-}6 \mu\text{g mL}^{-1}$	$0.82 \mu\text{g mL}^{-1}$ $0.28 \mu\text{g mL}^{-1}$ $0.02 \mu\text{g mL}^{-1}$ $0.49 \mu\text{g mL}^{-1}$	-	A.S	(13)
17	Carbamazepine	Exhaled breath condensate	-	Spectrofluorimetry	$0.2\text{-}20 \mu\text{g mL}^{-1}$	$0.08 \mu\text{g mL}^{-1}$	$0.39\text{-}0.51 \mu\text{g mL}^{-1}$	E.R	(14)

18	Tobramycin	Exhaled breath condensate	-	Spectrophotometry	1-50 ng mL ⁻¹	0.5 ng mL ⁻¹	13.7-31.1 ng mL ⁻¹	E.R	(15)
19	Phenobarbital	Plasma	Protein precipitation	Spectrophotometry	1-50 µg mL ⁻¹	0.6 µg mL ⁻¹	19 – 39.6 µg mL ⁻¹	E.R	(16)
20	Mycophenolic acid	Plasma	Protein precipitation	Spectrofluorimetry	1 – 10 µg mL ⁻¹	-	-	A.SH	(17)
21	Indoxyl sulfate	Plasma	Salting-out assisted liquid-liquid extraction	Spectrofluorimetry	2.5 – 40 µg mL ⁻¹		18.4–28.9 µg mL ⁻¹	F.N	Submitt ed.
22	Cynaide	Whole blood	-	Spectrophotometry	-	<1 µg mL ⁻¹	-	A.SH	Submitt ed.
23	Folic acid	Plasma	Protein precipitation	Spectrofluorimetry	0.01–1.1 mg/L	0.003 mg/L	-	JS	(18)
24	Methotrexate	Plasma	Protein precipitation	Spectrofluorimetry	0.02-10 µg/mL	0.015 µg/mL	-	M.SH	(19)
25	Deferiprone	Plasma and urine	Protein precipitation	Spectrofluorimetry	0.072–13 mmol/L	0.022 and 0.014 mM	-	JS	(20)
26	Doxorubicin	PlasmaUrine CF	Protein precipitation	Electrochemical	4.3×10^{-8} – 3.5×10^{-6} M 2.6×10^{-8} – 3.5×10^{-6} M 8.6×10^{-7} – 13×10^{-6} M	4.9×10^{-9} M 4.3×10^{-9} M 1.4×10^{-8} M	-	JS	(21)
27	Doxorubicin	Plasma Whole blood	Protein precipitation	Electrochemical	6.9×10^{-8} – 1.08×10^{-6} M 1.03×10^{-7} – 3.45×10^{-6} M	6.9×10^{-8} M 1.03×10^{-7} M	-	JS	(22)
28	Doxorubicin	Plasma	Protein precipitation	Electrochemical	0.086 µM to 3.45 µM	12 mM	-	PA	(23)
29	Doxorubicin	Plasma	-	Electrochemical	17 nM - 8.6 µM	17 nM	-	ME	(24)
30	Doxorubicin	Plasma	-	Electrochemical	0.018–3.6 µM	0.016 µM	-	N.H	(25)

31	Atenolol Carvedilol Propranolol	Serum	-	Electrochemical	12–96 µM 5–37 µM 10–104 µM	1.12 µM 3.01 µM 2.91 µM	-	N.S	(26)
32	Captopril	Serum	-	Electrochemical	0.06–0.2 µM	0.03 µM	-	M.H	(27)
33	Verapamil	Serum	-	Electrochemical	50–160 160–350 nmol dm ⁻³	41 nmol dm ⁻³	-	M.H	(28)
34	Diltiazem timolol	Serum	-	Electrochemical	0.1 to 100 µM 0.2 to 340 µM	0.06 µM 0.02 µM	-	M.H	(29)
35	Furosemide	Serum	-	Electrochemical	15–340 µM 17–100 µM	0.10 µM	-	M.H	(30)
36	Mefenamic acid indomethacin	Serum	-	Electrochemical	0.02–150 µM 0.08–435 M µM	0.02 µM 0.08 µM	-	M.H	(31)
37	Norfloxacin lomefloxacin	Serum	-	Electrochemical	0.06 to 170 µM 0.08 to 200 µM	0.2 µM 0.38 µM	-	M.H	(32)
38	Alprazolam Diazepam Clonazepam Oxazepam Chlordiazepoxide	Serum	-	Electrochemical	0.031-0.052 µM 0.027-0.041 µM 0.025-0.90 µM 0.025-0.047 µM 0.03-0.071 µM	0.031 µM 0.027 µM 0.025 µM 0.025 µM 0.03 µM	-	H.A	(33)
39	ketoconazole	Plasma	-	Spectrophotometry	10-80 µmol/L	10 µmol/L (LLOQ)	-	P.P	(34)

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References:

1. Zamani-Kalajahi M, Fazeli-Bakhtiyari R, Amiri M, Golmohammadi A, Afrasiabi A, Khoubnasabjafari M, et al. Dispersive liquid–liquid microextraction based on solidification of floating organic droplet followed by spectrofluorimetry for determination of carvedilol in human plasma. *Bioanalysis*. 2013;5(4):437-48.
2. Jouyban A, Sorouraddin MH, Farajzadeh MA, Somi MH, Fazeli-Bakhtiyari R. Determination of five antiarrhythmic drugs in human plasma by dispersive liquid–liquid microextraction and high-performance liquid chromatography. *Talanta*. 2015;134:681-9.
3. Jouyban A, Sorouraddin MH, Farajzadeh MA, Somi MH, Fazeli-Bakhtiyari R. Vortex-assisted liquid–liquid extraction combined with field-amplified sample injection and sweeping micellar electrokinetic chromatography for improved determination of β -blockers in human urine. *Talanta*. 2016;149:298-309.
4. Fazeli-Bakhtiyari R, Sorouraddin MH, Farajzadeh MA, Somi MH, Jouyban A. Salt-assisted LLE combined with field-amplified sample stacking in CE for improved determination of beta blocker drugs in human urine. *Bioanalysis*. 2014;6(3):319-34.
5. Alvani-Alamdari S, Jouyban A, Khoubnasabjafari M, Nokhodchi A, Rahimpour E. Efficiency comparison of nylon-6-based solid-phase and stir bar sorptive extractors for carbamazepine extraction. *Bioanalysis*. 2019;11(09):899-911.
6. Pourkarim F, Rahimpour E, Khoubnasabjafari M, Jouyban-Gharamaleki V, Jouyban A. Direct monitoring of verapamil level in exhaled breath condensate samples. *Pharm Sci*. 2019;25:50-6.
7. Feriduni B, Farajzadeh MA, Jouyban A. Determination Of Two Antiepileptic Drugs In Urine By Homogenous Liquid-Liquid Extraction Performed In A Narrow Tube Combined With Dispersive Liquid-Liquid Microextraction Followed By Gas Chromatography-Flame Ionization Detection. *Iranian journal of pharmaceutical research: IJPR*. 2019;18(2):620.
8. Pourkarim F, Rahimpour E, Khoubnasabjafari M, Jouyban-Gharamaleki V, Farhang S, Jouyban A. A Simple Colorimetric Method for Determination of Ethanol in Exhaled Breath Condensate. *Pharmaceutical Sciences*. 2020.
9. Jouyban A, Samadi A, Khoubnasabjafari M, Jouyban-Gharamaleki V, Ranjbar F. Amidosulfonic acid-capped silver nanoparticles for the spectrophotometric determination of lamotrigine in exhaled breath condensate. *Microchimica Acta*. 2017;184(8):2991-8.
10. Khoubnasabjafari M, Samadi A, Jouyban A. In-situ microscale spectrophotometric determination of phenytoin by using branched gold nanoparticles. *Microchimica Acta*. 2019;186(7):422.
11. Khoubnasabjafari M, Salari R, Samadi A, Jouyban-Gharamaleki V, Jouyban A. Colorimetric determination of phenytoin using indoxylo sulfate capped silver nanoparticles. *Analytical Methods*. 2019;11(26):3324-30.
12. Samadi A, Jouyban A, Amiraghiani N, Tayebi-Khosroshahi H. In situ Surfactant-based Solid Phase Microextraction of p-cresol in Human Plasma Prior to HPLC Analysis. *Current Analytical Chemistry*. 2020;16(6):687-94.
13. Samadi A, Khoubnasabjafari M, Barzegar M, Sadeghvand S, Shiva S, Jouyban A. Simultaneous Determination of Phenobarbital, Phenytoin, Carbamazepine and Carbamazepine-10, 11-epoxide in Plasma of Epileptic Patients. *Pharmaceutical Sciences*. 2019;25(4):345-51.
14. Hatifi A, Rahimpour E, Khoubnasabjafari M, Edalat M, Jouyban-Gharamaleki V, Alvani-Alamdari S, et al. A single-shot diagnostic platform based on copper nanoclusters coated with cetyl trimethylammonium bromide for determination of carbamazepine in exhaled breath condensate. *Microchimica Acta*. 2019;186(3):1-8.

15. Rezaei H, Rahimpour E, Khoubnasabjafari M, Jouyban-Gharamaleki V, Jouyban A. A colorimetric nanoprobe based on dynamic aggregation of SDS-capped silver nanoparticles for tobramycin determination in exhaled breath condensate. *Microchimica Acta*. 2020;187(3):1-9.
16. Zadaliasghar S, Rahimpour E, Khoubnasabjafari M, Pournaghi-Azar MH, Nokhodchi A, Jouyban A. A nano-platform for phenobarbital determination based on its inhibitory effect on the aggregation of silver nanoparticles/melamine system. *Journal of Molecular Liquids*. 2020;316:113891.
17. Shayanfar A, Ghavimi H, Zolali E, Jouyban A. Determination of Mycophenolic Acid in Plasma Samples Using the Terbium-Sensitized Luminescence Method. *Journal of Applied Spectroscopy*. 2015;82(4):614-9.
18. Manzoori JL, Jouyban A, Amjadi M, Soleymani J. Spectrofluorimetric determination of folic acid in tablets and urine samples using 1, 10-phenanthroline-terbium probe. *Luminescence*. 2011;26(2):106-11.
19. Jouyban A, Shaghaghi M, Manzoori JL, Soleymani J, JalilVaez-Gharamaleki J. Determination of methotrexate in biological fluids and a parenteral injection using terbium-sensitized method. *Iranian journal of pharmaceutical research: IJPR*. 2011;10(4):695.
20. Manzoori JL, Amjadi M, Soleymani J, Tamizi E, Rezamand A, Jouyban A. Determination of deferiprone in urine and serum using a terbium-sensitized luminescence method. *Luminescence*. 2012;27(4):268-73.
21. Soleymani J, Hasanzadeh M, Shadjou N, Jafari MK, Gharamaleki JV, Yadollahi M, et al. A new kinetic-mechanistic approach to elucidate electrooxidation of doxorubicin hydrochloride in unprocessed human fluids using magnetic graphene based nanocomposite modified glassy carbon electrode. *Materials science and engineering: C*. 2016;61:638-50.
22. Soleymani J, Hasanzadeh M, Eskandani M, Khoubnasabjafari M, Shadjou N, Jouyban A. Electrochemical sensing of doxorubicin in unprocessed whole blood, cell lysate, and human plasma samples using thin film of poly-arginine modified glassy carbon electrode. *Materials science and engineering: C*. 2017;77:790-802.
23. Alizadeh PM, Hasanzadeh M, Soleymani J, Gharamaleki JV, Jouyban A. Application of bioactive cyclic oligosaccharide on the detection of doxorubicin hydrochloride in unprocessed human plasma sample: A new platform towards efficient chemotherapy. *Microchemical Journal*. 2019;145:450-5.
24. Ehsani M, Soleymani J, Hasanzadeh M, Vaez-Gharamaleki Y, Khoubnasabjafari M, Jouyban A. Sensitive monitoring of doxorubicin in plasma of patients, MDA-MB-231 and 4T1 cell lysates using electroanalysis method. *Journal of Pharmaceutical and Biomedical Analysis*. 2020;113701.
25. Hashemzadeh N, Hasanzadeh M, Shadjou N, Eivazi-Ziae J, Khoubnasabjafari M, Jouyban A. Graphene quantum dot modified glassy carbon electrode for the determination of doxorubicin hydrochloride in human plasma. *Journal of pharmaceutical analysis*. 2016;6(4):235-41.
26. Shadjou N, Hasanzadeh M, Saghatforoush L, Mehdizadeh R, Jouyban A. Electrochemical behavior of atenolol, carvedilol and propranolol on copper-oxide nanoparticles. *Electrochimica acta*. 2011;58:336-47.
27. Hasanzadeh M, Pournaghi-Azar MH, Shadjou N, Jouyban A. Electropolymerization of taurine on gold surface and its sensory application for determination of captopril in undiluted human serum. *Materials Science and Engineering: C*. 2014;38:197-205.
28. Hasanzadeh M, Pournaghi-Azar MH, Shadjou N, Jouyban A. A verapamil electrochemical sensor based on magnetic mobile crystalline material-41 grafted by sulfonic acid. *Electrochimica Acta*. 2013;89:660-8.
29. Hasanzadeh M, Pournaghi-Azar MH, Shadjou N, Jouyban A. Determination of diltiazem in the presence of timolol in human serum samples using a nanoFe 3 O 4@ GO modified glassy carbon electrode. *RSC Advances*. 2014;4(93):51734-44.
30. Hasanzadeh M, Pournaghi-Azar MH, Shadjou N, Jouyban A. A new mechanistic approach to elucidate furosemide electrooxidation on magnetic nanoparticles loaded on graphene oxide modified glassy carbon electrode. *RSC Advances*. 2014;4(13):6580-90.

31. Hasanzadeh M, Shadjou N, Saghatforoush L, Dolatabadi JEN. Preparation of a new electrochemical sensor based on iron (III) complexes modified carbon paste electrode for simultaneous determination of mefenamic acid and indomethacin. *Colloids and Surfaces B: Biointerfaces*. 2012;92:91-7.
32. Saghatforoush LA, Sanati S, Mehdizadeh R, Hasanzadeh M. Solvothermal synthesis of Cd (OH) 2 and CdO nanocrystals and application as a new electrochemical sensor for simultaneous determination of norfloxacin and lomefloxacin. *Superlattices and Microstructures*. 2012;52(4):885-93.
33. Ashrafi H, Mobed A, Hasanzadeh M, Babaie P, Ansarin K, Jouyban A. Monitoring of five benzodiazepines using a novel polymeric interface prepared by layer by layer strategy. *Microchemical Journal*. 2019;146:121-5.
34. Pashazadeh-Panahi P, Hasanzadeh M, Eivazzadeh-Keihan R. Spectrophotometric study of ketoconazole binding with citrate capped silver nanoparticles and its monitoring in human plasma samples. *Journal of Molecular Recognition*. 2020;33(5):e2830.

